



Effect of local sea cucumber (*Phyllophorus* sp.) methanol extract as natural antifouling against macrofouler (*Nerita* sp.)

Himatul Hasanah*, Rizqi Abdi Perdanawati, and Dian Sari Maisaroh

Marine Science Department, Faculty of Science and Technology, UIN Sunan Ampel Surabaya, Indonesia

*E-mail: hhima.hs20@gmail.com

ABSTRACT

Macrofouling is the activity of attaching biota with size > 0.5 cm, which attach to and form a colony in the mariculture construction. The development of antifouling materials currently uses a lot of natural ingredients including marine life, namely sea cucumbers. This study aims to determine the activity of local sea cucumber (*Phyllophorus* sp.) Methanol extract as a natural antifouling against macrofouler *Nerita* sp. The experimental design for this study used a completely randomized design with five types of treatment, namely negative control treatment, extract concentration of 75 mg/ml, extracts concentration of 100 mg/ml, extracts concentration of 200 mg/ml and positive control with three repetitions for each treatment. The results showed that the local sea cucumber (*Phyllophorus* sp.) methanol extract contained saponins, alkaloids, tannins, steroids and phenolic bioactive compounds. The local sea cucumber (*Phyllophorus* sp.) methanol extract has an effect on the antifouling rate value. The same thing also happened where the increasing concentration of local sea cucumber (*Phyllophorus* sp.) methanol extract gave a significant decrease in the regaining ability of macrofouler *Nerita* sp. Lethal concentration (LC_{50}) values of local sea cucumber methanol extract (*Phyllophorus* sp.) on macrofouler *Nerita* sp. were investigated. While the lethal concentration (LC_{50}) value of the lethal dose was estimated at 185.18 mg/ml. This study shows that local sea cucumber (*Phyllophorus* sp.) has potential as an antifouling agent.

Keywords: sea cucumber, *Phyllophorus* sp., antifouling, macrofouler, *Nerita* sp.

INTRODUCTION

The existence of macrofouling becomes one of the serious problems caused by the risk of losses it causes. Macrofouling is the activity of attaching biota that has a size of >0.5 cm which lives on and forms a colony [1]; [2]. Macrofouling has the ability to grow and develop rapidly in various constructions submerged by water. Accumulation process macrofouling that occurs sustainable development could raises problems both economically and operations [3]. Macrofouling consists of various groups include algae, mollusks, crustaceans, bryozoa and polychaeta. One type macrofouling that is thought most often found in coastal waters in jember area the mollusks, *Nerita* sp.

Nerita sp. is a type of mollusk gastropods that have a habit of life clings to rocky beaches as well as Breakwater. They include animals that soft-bodied and covered with a shell [4]. *Nerita* sp. can be herbivorous, carnivorous, omnivorous or detritivores [5]. The existence of mollusks such as *Nerita* sp. allegedly resulting in impacts and losses for the environment. One of the impacts is its existence on a cultivation raft which causes the raft to sink [6]; [7]. In addition, the attachment of these biota too cause the occurrence damage to the floating bag net and an effect on decreasing seaweed production [8]. The presence of molluscs could causing net closure in aquaculture so as to increase the mortality of cultured fish and result in the spread of disease [9]; [10]; [11]; [12]. Embedding an

organism on metal materials can intensively corrosion [13]. Damage to wood in coastal structures or ships occurs due to the attack of the attaching biota which causes surface changes [14]; [15]. Repair efforts because the damage was caused existence macrofouling requires a large amount of money as explained by [16] which states that the government and industry in America spent at least 200 million dollars to solve the problem. The damage brought about by existence macrofouling provide serious losses so that prevention efforts are needed. Prevention macrofouling usually done using antifouling paint.

Paint antifouling this prevent the occurrence macrofouling by creating an effective and constant biocide [7]. In general antifouling paint containing copper and TBT (tri-n-butyltin) as the active and most effective elements [7]. The existence of the active ingredients contained in the anti-fouling paint creates new problems because TBT is not only toxic to fouling organisms but also harmful to non-target organisms. In addition, TBT was also reported to be very bad poisonous and persistent in marine environment [17]; [18]. This is a new challenge for industry and government in developing alternative technologies to prevent pollution in construction and installation mariculture. Based on This problem requires an alternative compound that is environmentally friendly so that it does not cause disturbance and damage to non-target organisms or the marine environment. One of the alternative compounds that can be used is extracts from

marine life. Lately, many people use natural products as antifouling agents wrong only one with utilize marine life. a marine biota that can produce compounds antifouling s sea cucumber extract. Sea cucumber extract contains secondary metabolites such as alkaloids, steroids, sapogenin, saponins, triterpenoids, glycosaminoglycan, lectins, phenols and flavonoids [19];[20]. Based on the content of bioactive compounds it has, *Holothuria* can be used as antibacterial, antimicrobial, antifouling, anti-cancer, anti-tumor and immunostimulant [20]; [21].

Based on the facts that have been described, it is necessary to carry out further research regarding the effect of adding local sea cucumber methanol extract (*Phyllophorus* sp.) as antifouling an alternative to TBT compounds to make it more environmentally friendly. Laboratory tests are carried out to determine whether sea cucumber extract has the ability and potential as antifouling, so this research aims to find out whether addition extract methanol Local sea cucumbers can affect biota fouling.

RESEARCH METHOD

This research was conducted from January to July 2020. The materials used in this research are local sea cucumbers (*Phyllophorus* sp.) obtained from Kenjeran Beach, Surabaya. The test biota used is macrofouler *Nerita* sp. obtained from Pancer Beach, Jember.

This study used laboratory experimental methods. Collection Laboratory scale data is carried out by observing activities antifouling rate, regaining rate and lethal concentration 50. The experimental design for this study used a completely randomized design (CRD) with five types of treatment, namely negative control treatment, extract concentration of 75 mg/ml, extract concentration of 100 mg/ml, extract concentration of 200 mg/ml and positive control with three repetitions for each treatment.

Samples collection

Local sea cucumber samples (*Phyllophorus* sp.), obtained from sea cucumber fishermen at Kenjeran Beach, Surabaya (Fig. 1). The sea cucumber samples were stored in cooler box and given ice cubes with the aim of maintaining the freshness of sea cucumbers. Then the sea cucumber samples are taken to the laboratory for the extraction and identification process.

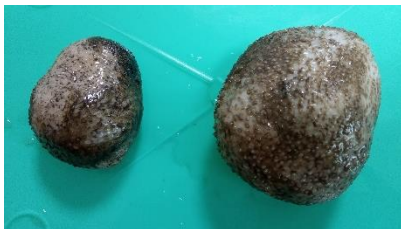


Fig. 1 Sample of local sea cucumber (*Phyllophorus* sp.)

The test biota used in this study is macrofouler that is *Nerita* sp. *Nerita* sp. used has a weight 0.65 gram. Operculum brownish in color, chalky like a thick plate, has hooks and a grained outer surface. Black shell with spots brownish. *Nerita* sp. is a group herbivorous animal. The test biota was obtained from coral and break water at Pancer Puger Beach, Jember (Fig. 2).



Fig. 2 *Nerita* sp.

Acclimatization Process

The acclimatization process is an effort to adjust the physiological or adaptation of an organism to a new environment that it will enter [22]. After 3 days of acclimatization, the local sea cucumber were used for the antifouling test.

Extraction method

The extraction method in this study used maceration (immersion) with use solvent polar [23]. The polar solvent used in this study is a methanol solvent. Sample sea cucumber cleaned with Secrete all over stomach sea cucumber then washed with water flows. Cut sea cucumber small with purpose to enlarge the contact surface area with ingredient. sea cucumber Soaked in methanol solvent at room temperature with a ratio of mass and volume of extract making 1: 4 means that 1 gram of sea cucumber in 4 ml of methanol solvent (Fig. 3).

Maserat stored in a container closed and not exposed to light. The maserate is then filtered, the filtrate is separated and the pulp is soaked back into the new solvent. The maceration process is carried out up to obtained a clear filtrate, which means that the solution is saturated. The filtrate obtained is concentrated through a simple distillation process. The result obtained is a paste later made three sorts concentrations with distilled water were 75 mg/ml, 100 mg/ml and 200 mg/ml.



Fig. 3 Local sea cucumber methanol extract

Bioactive Compound Test

The extraction results obtained were analyzed qualitatively to determine womb compound bioactive in sea cucumbers. Qualitative analysis using phytochemical screening as follows:

Alkaloid Test method Culvenor-Fitzgerald

A sample of 1 ml is mixed with 1 ml of chloroform and 1 ml of ammonia is put into a test tube then heated over a water bath, shaken and then filtered. The filtrate obtained is divided into three equal parts and then put into the tube each filtrate was taken and tested using mayer, wagner and dragendorf reagents. The

formation orange, brown, and white deposits on each test indicating the presence of alkaloids [24].

Flavonoid Test

A sample of 1 ml was mixed with 70% ethanol and then shaken and heated, shaken again then filtered. The filtrate obtained then Mg 0.1 g and 2 drops of concentrated HCl are added. The presence of flavonoids is characterized by their formation the red color on the ethanol layer.

Saponin Test

Saponin test is carried out by means of a sample of 1 ml boiled in 10 ml of water in a water bath. The filtrate was shaken and let stand for 15 minutes. Existence compound saponins indicated by the formation stable foam.

Steroid Test

A sample of 1 ml was mixed with 3 ml of 70% ethanol and added 2 ml of H₂SO₄ concentrated and 2 ml anhydrous acetic acid (reagent Libermann-burchard). The presence of steroids is indicated by a color change from purple to blue / green.

Triterpenoid

Test compound triterpenoids done with add 1 ml of sample to 2 ml of chloroform and 3 ml of acid sulfate concentrated. Existence triterpenoid compounds are characterized with formation color red brownish between surface.

Phenolic Test

Sample as much 1 ml put into the test tube then add 1 ml of 1% NaCl solution and 1 ml of solution gelatin 10 %. The formation of color deposits outih showing existence phenolic compounds.

Tannin Test

A sample of 1 ml is boiled in 20 ml of water over a water bath after which it is filtered. Filtrate that obtained add 2-3 drops of 1% FeCl₃. The presence of tannin compounds is indicated by formation color greenish brown or blackish blue.

Water Quality Measurement

Based on [18] growth influenced by the presence of abiotic components which include the physical properties of seawater such as salinity, brightness, pH, temperature, tides and currents. Water quality parameters measured in this study include salinity, temperature, pH and DO.

Antifouling Test

Method that used in this research is an anti-sticking method. This method was chosen because it is considered to be faster in obtaining results. The antifouling test process begins with preparing a 500 ml aquarium containing sea water containing local sea cucumber methanol extract (*Phyllophorus* sp.). *Nerita* sp. which has acclimatized transferred to a different aquarium. total *Nerita* sp. that used is 10 fish for each aquarium. The test was carried out with three replications for each treatment. Observation *Nerita* sp. carried out every 1 × 24 hours and 2 × 24 hours after exposure. The analysis used in this

study includes anti-adhesion analysis, regaining rate and analysis lethal concentration 50 or LC₅₀.

This study used a factorial completely randomized design method.

A factorial completely randomized design is a design with two or more factors in an environment that is considered homogeneous [25]. The factor used in the study was the concentration of sea cucumber extract which was thought to contain antifouling compounds. The concentration factor of sea cucumber extract has three levels, namely 75 mg / ml, 100 mg / ml, 200 mg / ml concentration factor extract sea cucumber presumed effect on the observed response, namely the number of attached biota, regaining rate. The hypothesis is tested with using the Anova test using software SPSS.

Score LC₅₀ obtained with use probit method so that obtained a linear regression equation $y = a + bx$. Price y stated that 50% of the test biota died for 2 × 24 hours. The value of a and b values are the slope and intercept values of the three concentrations used. The value of x obtained is the concentration solution that cause mortality in the test biota by 50%

$$Y = ax + b \quad (1)$$

Price y stated that 50% of the test biota died for 48 hours. The value of a and b values are the slope and intercept values of the three concentrations used. The value of x obtained is the concentration solution that cause mortality in the test biota by 50%.

RESULT AND DISCUSSION

Phytochemical test of local sea cucumber methanol extract (*Phyllophorus* sp.)

The phytochemical test is a qualitative test for the content of bioactive compounds (secondary metabolites) contained in the sample. The bioactive compounds tested include alkaloids, flavonoids, saponins, triterpenoids, steroids, phenolics and tannins. This is done aim at to knowing further benefits of the bioactive compounds produced. Extract analysis results local sea cucumber methanol (*Phyllophorus* sp.) are presented in **Table 1**. Based on the table, it is known that methanol extract local sea cucumber (*Phyllophorus* sp.) are known to have compound metabolites secondary including alkaloids, phenolics, steroids, saponins and tannins.

Table 1 The result of phytochemical content screening of *Phyllophorus* sp.

Target	The Result
Alkaloid	+
Phenolic	+
Steroid	+
Triterpenoid	A presence of brownish red
Flavonoid	A formation of reddish colour
Saponin	+
Tannin	+

(+ indicates the presence of the substance)

Positive reaction from the alkaloid test if the solution is formed an orange precipitate, a brown precipitate and sediment white after added reagent mayers, wagner and dragendorff. Alkaloids are compounds organic which is basic and has quite a lot of hydrogen atomic groups. The hydrogen atom group in alkaloids can cause anesthetic effects and even death in animals depending on the dose given [3]. Alkaloid compounds produced by ascidians can reduce biofouling growth especially barnacles [26]. Alkaloids have the ability to be antibacterial because they can inhibit them process formation peptidoglycan in bacterial cells so that the cell wall layer is not formed completely, causing cell death [27]; [28].

Test phenolic done with adding 10% NaCl and 1% gelatin solution. The presence of phenolic compounds is indicated by the formation of a white precipitate. Phenolic compounds are compounds with a high acidity when compared to alcohol. Phenol can cause chemical burning on the skin, the high level of phenol acidity can cause death in biofouling [3]. The phenol group has the ability to damage cell membranes, activate enzymes as well denature protein that cause cell wall damage due to subsidence ability permeability. Changes in cytoplasmic permeability can disrupt the transport of important organic ionions into cells so that they can inhibit growth and cell death [29].

Results of identification steroid compounds using reagents Libermanburchard in local sea cucumber methanol extract (*Phyllophorus* sp.) showed positive results. A positive reaction to the steroid test is a color change from purple to blue or green. Steroids are triterpenoid class compounds and are usually used as the basis for making drugs. Extracts containing steroid compounds have potential as antibacterial and anti-fungal [30]. Steroids have the ability to inhibit bacterial growth by damaging the bacterial cell membrane. This is also explained by [31] which states that the mechanism of steroids as antibacterial is closely related to membrane lipids and sensitivity to components steroids that cause leaks in the liposomes. Steroids can interact with cell membrane phospholipids which are permeable to lipophilic compounds so that could cause drop integrity membrane and changes in cell membrane morphology causing cells to become brittle and lysis [32].

Tannin compounds identified in local sea cucumber methanol extract (*Phyllophorus* sp.). The positive reaction of tannin compounds after adding FeCl_3 1% is formed a brown color. Tannins are a compound polyphenols that form insoluble complex compounds with protein. Tannins have ability to inhibit the activity of several digestive enzymes such as triptin, chymotrypsin amylase, and lipase besides that it can also inhibit iron absorption [33]. Tannins have ability inhibits bacterial growth by using the peptidoglycan synthesis process which results in the formation of imperfect cells. Cell wall shrinks and permeability disturbed so that inhibits bacterial growth [23]. Based on the statement of [3] tannin compounds have an OH group which functions to reduce the oxidation and reduction reaction processes so that they have the ability to inhibit the release of $\text{Fe} + 2$ ions as a cause of corrosion and the attachment of fouling biota to the surface layer of the iron plate.

Saponins are produced as a form of chemical self-defense for sea cucumbers in nature. Saponins are also believed to have biological effects including anti-fungal, cytotoxic against tumor cells, hemolysis, immunostimulants and anti-cancer [34]. The main effect of saponins on bacteria is the release of proteins and enzymes in cells [35]. Saponins function as antimicrobials (antifouling) by inhibiting or killing microbes by interacting with sterol inhibitors [36];[37]. Saponins in sea cucumbers have been characterized as holothurin compounds [26]. Saponins have also been reported as toxic or bioactive compounds [38].

Antifouling test of local sea cucumber (*Phyllophorus* sp.) methanol extract against *Nerita* sp.

In order to determine the ability of local sea cucumber methanol extract (*Phyllophorus* sp.). In inhibiting the attachment of biofouling, a measurement parameter is needed that can represent the relationship, one of which is a parameter antifouling rate. Analysis antifouling rate used to determine the level of adherence (inhibition) of local sea cucumber methanol extract (*Phyllophorus* sp.) against macrofouler *Nerita* sp. Before conducted antifouling test, it is necessary to check the condition of the maintenance water quality first with purpose to ensure that the water quality is in accordance with the quality standards required by the biota *Nerita* sp. Test the water quality 3 repetitions for each parameter. The purpose of measuring water quality in this study is to control the conditions of water quality during maintenance. The measured water quality test includes temperature, salinity, DO and pH. Measurement of water quality is used as supporting data to support it research this. Result from water quality measurement are presented in **Table 2**.

Table 2 Water Quality Measurement

Parameter	Mean	Optimum Value	Source
Salinity	31,2 °C	28 – 32	KEPMEN.LH No.59 Year 2004
Temperature	34 ppt	33 – 34	KEPMEN.LH No.59 Year 2004
DO	7,6	7 – 8,5	KEPMEN.LH No.59 Year 2004
Ph	3 mg/ml	3 – 7	Kadim, et al., 2017

Temperature is one of the parameters that plays an important role for the survival of marine life both in situ and exitu. Temperature affects the rate of metabolism of aquatic ecosystems. Measurement results temperature in research showing score amounting to 31.2 °C. Based on [39] the optimal temperature value for marine biota ranges from 28-32 °C.

Salinity the presence of adhering biota in the waters, this is because each organism has a different tolerance to salinity. The results of the salinity measurement in the study showed a result of 34 ppt. Based on [39], the optimal salinity value for biota ranges from 33-34 ppt. Tthe salinity for the growth of adhering biota ranges from 30-33 ppt.

Based on the measurement results, the pH value in the study showed a value of 7.6. Based on [29], the

optimal pH for marine life ranges from 7 - 8.5, which means that the pH value in the study is classified as optimal for the maintenance of adhering biota. The degree of acidity, also known as pH, has a less significant direct effect on the attachment of biota fouling. This has been reported by [40] which states that a decrease in pH has a less significant direct effect on larval development and attachment of fouling biota.

Dissolved oxygen is one of the important parameters needed by biota for the respiration process and the breakdown of organic substances. The need for dissolved oxygen tends to vary depending on the type, stage and activity [41]. The results of measuring DO at the time of the study were obtained a value of 3 mg / l. The optimal DO value for marine life ranges from 3 - 7 mg/l [42].

Antifouling Rate

So that knowing ability local sea cucumber methanol extract (*Phyllophorus* sp.) in hinder sticking biofouling, a measurement parameter that can represent the relationship is needed, one of which is the antifouling rate parameter. Analysis antifouling rate use to know level sticking (inhibition) local sea cucumber methanol extract (*Phyllophorus* sp.) against macrofouler *Nerita* sp. (Fig. 4).

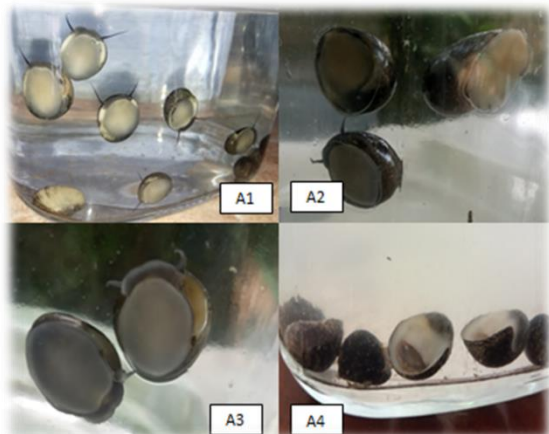


Fig. 4 Type of Treatment (A1) Treatment of negative control, (A2) Treatment of sea cucumber extract concentration of 75mg/ml, (A3) Treatment of sea cucumber extract concentration of 100mg/ml, (A4) Treatment of sea cucumber extract concentration of 200mg/ml.

The results of the local sea cucumber extract antifouling activity test (*Phyllophorus* sp.) on a laboratory scale in A1 treatment, namely negative control, it shows that all test biota is attached to the aquarium wall while the opposite occurs in treatment A5, namely positive control where almost all test biota does not stick to the aquarium wall. This happens because in the A5 treatment an antifouling paint is added so that the chemical compounds contained in the paint are exposed to water so that it affects the physiological conditions of the macrofouler. *Nerita* sp. which causes inhibition due to the antifouling compound of the paint.

Based on the results of the antifouling test, it shows that some biota is still attached *Nerita* sp (Fig. 4) on the aquarium. Condition *Nerita* sp. that is not attached is indicated by closing tightly operculum and is at the bottom

of the aquarium (Fig. 5). In this research, when an active liquid stimulus such as methanol extract from local sea cucumber was exposed on the foot, *Nerita* sp. contracted its foot immediately.



Fig. 5 The condition of *Nerita* sp. at the bottom of the aquarium

Percentage value of antifouling for 1 x 24 hour observation in A1 treatment (negative control), A2 treatment (concentration 75 mg / ml), A3 treatment (100 mg / ml), A4 treatment (200 mg / ml) and A5 treatment (positive control) at 0%, 50%, 53.3%, 80% and 90%, respectively. The percentage of antifouling for observation 2 x 24 hours in treatment A1 (negative control), treatment A2 (concentration 75 mg / ml), treatment A3 (100 mg / ml), treatment A4 (200 mg / ml) and treatment A5 (positive control) respectively 0%, 50%, 60%, 73% and 87% (Fig. 6)

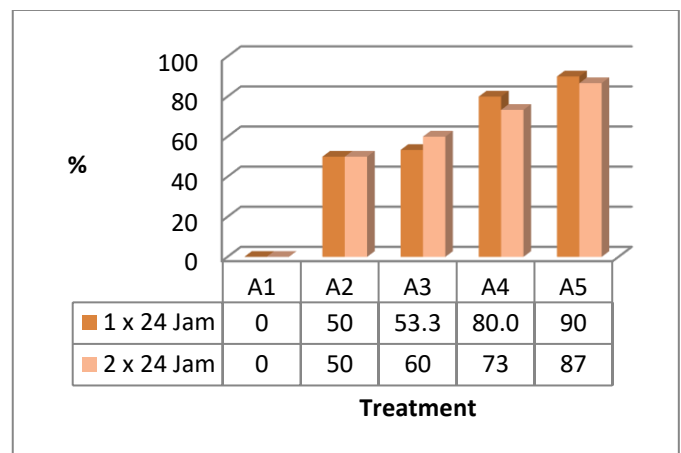


Fig. 6 Antifouling rate

It is known that the higher the extract concentration, the greater the antifouling rate value (Fig. 6). This is thought to be due to the fact that the higher the extract concentration, the higher the saponins, alkaloids, steroids, phenolics and tannins. Womb the have ability as an antifouling. This is in accordance with research from [43], in the study it was known that tannins could hinder growth biofouling even at low concentrations.

The results of this study indicate that *Nerita* sp. disturbed with addition of local sea cucumber methanol extract (*Phyllophorus* sp.). This is due to the local sea cucumber methanol extract (*Phyllophorus* sp.) contains secondary metabolite compounds which function as antifouling. Sea cucumbers contain many toxic compounds such as saponins (Holothurin). These toxic compounds can be used as a source of potent antifouling [44].

Alkaloid and phenolic compounds also reduce could total sticking biofouling because these two

compounds are poisonous which can kill biofouling [30]. To knowing influence the concentration of local sea cucumber methanol extract (*Phyllophorus* sp.) to the antifouling rate it is necessary to test One Way ANOVA. Before the test One Way ANOVA is done so first it is necessary to test for normality and homogeneity.

Regaining Rate

Other factors besides the antifouling rate can be used to determine whether the local sea cucumber methanol extract (*Phyllophorus* sp.) can be used as an antifouling is the regaining rate. The test biota that has been tested for antifouling is then transferred to the aquarium for analysis regaining rate. Regaining rate analysis was performed to look ability recovery of the test biota after receiving treatment from addition extract local sea cucumber methanol (*Phyllophorus* sp.). (Table 3). The results of the regaining rate analysis show the results as in the following figure (Fig. 7).

Based on picture indicates that the value regaining rate in each different treatment. The regaining rate values for treatment A1, A2, A3, A4 and A5 were 96.7%, 85%, 60%, 45% and 6.7%, respectively.

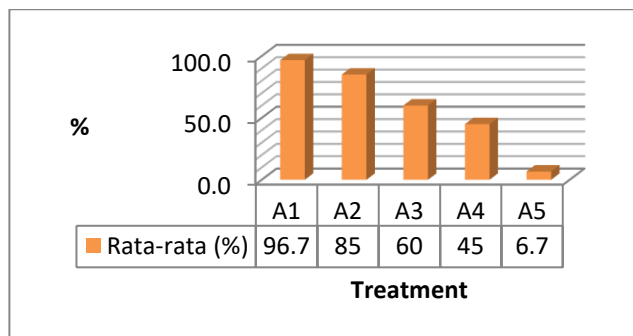


Fig. 7 Regaining rate

This value shows that the higher the concentration of local sea cucumber methanol extract (*Phyllophorus* sp.) is given then the value regaining rate the smaller it is, this is because the bioactive compounds in the extract exceed the tolerable limit *Nerita* sp so that it affects his recovery ability.

Table 3 Behavior of the test biota after being treated in fresh sea water

Concentration	Behavioural change of snails observed during exposure and after in fresh sea water
Control (-)	Open operculum and rapid movement
75 mg/ml	Moving with spread foot and regained immediately
100 mg/ml	Start moving within 10 minutes
200 mg/ml	Start removing the foot within 30 minutes and part of the test biota died
Control (+)	Almost all of the test biota died, <i>Nerita</i> sp. floats on the surface of the water with body part popping out of the shell

Behavioural change of *Nerita* sp. observed during exposure and after in fresh sea water, it showed a different response in each treatment. Response *Nerita* sp. at a concentration of 75 mg / ml shows a response in the form

of a snail starting to remove its antennae and walking quickly. Response *Nerita* sp. at a concentration of 100 mg/ml is characterized by its onset movement after 10 minute put into sea water. A concentration of 200 mg / ml shows a response such as starting removing foot within 30 minutes, some of the test biota died. Provision of local sea cucumber methanol extract (*Phyllophorus* sp.) causes a reaction between the active substance and the receptors in the effector organ causing symptoms of poisoning. Each test biota used gives a different response at a certain concentration. The difference in response is due to differences in the sensitivity level of each biota (Fig. 8).

Influence Antofouling Rate To Regaining Rate

Based statistical test between concentrations local sea cucumber methanol extract (*Phyllophorus* sp.) against the antifouling rate macrofouler *Nerita* sp. It is known that the extract concentration has a significant effect on the antifouling rate macrofouler *Nerita* sp. The same thing happened to the results of the analysis between the extract concentration and the regaining rate, which also showed a significant influence between the extract concentration and the regaining rate. macrofouler *Nerita* sp.

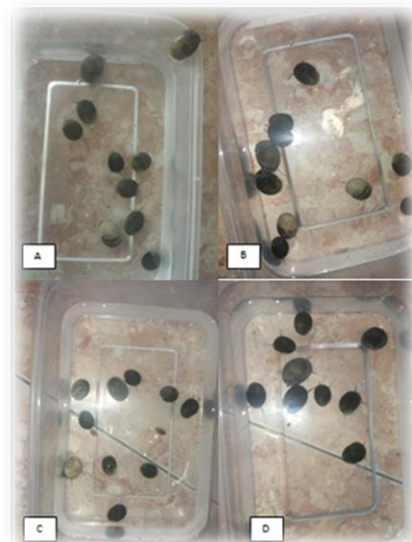


Fig. 8 (A) A1 treatment, (B) A2 treatment, (C) A3 treatment, (D) A4 treatment

Follows up on the above, giving rise to the question, whether the antifouling rate and regaining rate have a correlation with each other. To find out this, it is necessary to do further tests. One of the tests that can be done is a simple linear regression test. Simple linear regression analysis is used to determine whether the antifouling rate and regaining rate factors influence each other.

Analysis Lethal Concentration 50 (LC₅₀)

Rate the percentage of deaths macrofouler *Nerita* sp. at each concentration showed different results. The largest percentage value of death occurred in treatment A5 which was 90%, then followed by A4 treatment which was 50%. The smallest percentage value of death in treatment A1 is 0%. This mortality percentage value is then used to perform LC₅₀ by using the probit method [45].

LC₅₀ carried out with the aim of knowing the concentration that can cause death by 50% of the test organism which can be estimated using graphs and calculations on a observation certain. Lethal concentration 50 or LC₅₀ is a calculation to determine the activity of an extract or compound. LC₅₀ 48H can be determined using the test statistics (Fig. 9).

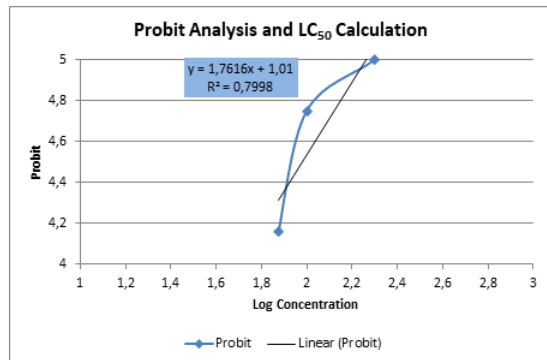


Fig. 9 Probit analysis and LC₅₀ calculations

Based on the calculation results lethal concentration 50 or LC₅₀ earned value amounting to 185.18. This means concentration sea cucumber extract (*Phyllophorus* sp.) is lethal in 50% of all tested biota, namely at a concentration of 185.18 mg / ml. Score lethal concentration 50 or LC₅₀ amounted to 185.18 based on the low toxicity category [46].

Local sea cucumber methanol extract (*Phyllophorus* sp.) has the ability to kill fouling biota even in low concentrations. The total concentration of local sea

CONCLUSIONS

Local sea cucumber methanol extract (*Phyllophorus* sp.) have content of compounds such as saponins, alkaloids, tannins, steroids and phenolics. It give influence on the value of the antifouling rate by reducing the adhesion rate macrofouler *Nerita* sp. with increasingly increasing concentration. The same thing happened Where increasingly increase in extract concentration give drop significant on regaining ability macrofouler *Nerita* sp. It also has an impact on value nerita's macrofouler regaining rate sp. where the smaller the concentration given, the regaining rate value macrofouler *Nerita* sp. the greater it is. Variation concentration that applied to give a response different against macrofouler *Nerita* sp.

REFERENCES

- [1] Baveridge, C.M.1987. Cage Aquaculture. Dorset Press. Porchester. 365 pp
- [2] Rejeki, S. 2009.Sukses Penempelan Makro Marine-biofouling Pada Jaring Karamba Apung di Teluk Hurun Lampung. Ilmu Kelautan. vol. 14 (2) : 112-117
- [3] Syahputra, F., & Almuqaramah, T. 2019. Penambahan ekstrak larutan kulit mangrove pada cat minyak sebagai antifouling. Aquatic Sciences Journal, 37-40.Tambaru, R. 2000. Pengaruh Waktu Inkubasi Terhadap Produktivitas Primer Di Perairan Teluk Hurun. (Thesis). Bogor. Program Pasca Sarjana; Insitut Pertanian Bogor
- [4] Basri, N.B, Syawal N.H, & Baharuddin N. 2018. Marine gastropods (Gastropoda; Mollusca) diversity and distribution on intertidal rocky shores of Terengganu, Peninsular Malaysia. AACL Bioflux , 1144-1154
- [5] Eichhorst, T. 2016. Neritidae Of The World. Conchbooks, 694 pp.
- [6] Armstrong, E., Boyd, K.G. & Burgess, J.G. 2000. Prevention of marine biofouling using natural compounds from marine organisms, Biotechnology Annual Review. Elsevier, pp. 221-241
- [7] Marhaeni, B. 2008. Biofouling Pada Beberapa Jenis Substrat Permukaan Kasar dan Halus. Sains Akuatik.

- [8] La Didu, Ma'ruf K., & Emiyarti. 2019. Komposisi Jenis dan Kepadatan Makrobiofouling Pada Jaring Kantung Apung Dengan dan Tanpa Menggunakan Sintetik Anti Fouling Hubungannya dengan Pertumbuhan *Kappapycus alvarezii* Di Perairan Pantai Lakeba Kota Baubau . Jurnal Manajemen Sumber Daya Perairan, 111-121
- [9] Huang, C.C. 2000. Engineering Risk Analyses for Sub-merged Cage Net System in Taiwan. In Cage Cul-ture in Asia: Proceeding for the First International Symposium on Cage Aquaculture in Asia (ed. IC. Liao and C.K. Lin), 133– 40 pp
- [10] Phillipi, A.L., N.J. O'Connor, A.F. Lewis, & Y.K. Kim, 2001. Surface Flocking as a Possible Anti-biofou-lant. Aquaculture 195: 225 – 238Pranoto, E., W.F
- [11] Tan, C.K.F., B.F., Nowak, & S.L.Hodson, 2002. Biofoul-ing as Reservoir of *Neoparamoeba pemaquiden-sis*, the Causive Agent of Amoebic Gill Disease in Atlantic Salmon. Aquaculture 210: 49-58
- [12] Swift, M.R., D.W.Fredericson, A. Unrein, B. Fullerton, O. Patursson, & K. Baldwin, 2006. Drag Force Act-ing on Biofouled Net panels. Aquaculture Engi-neering 35: 292-299
- [13] Railkin, A. 2004. Marine biofouling: Colonization processes and defenses. Boca Raton, FL, USA: CSC Press.
- [14] PEREIRA, R.C., M. D. PINHEIRO and B.A.P. DA GAM A .2002. Feeding preference of the endemic gastropod *Astraea latispina* in relation to chemical defenses of Brazilian tropical seaweeds. Braz. J. Biol., 62 : 33-40
- [15] Boesono, H. 2008. Pengaruh Lama Perendaman Terhadap Organisme Penempel dan Modulus Elastisitas Pada Kayu. Ilmu Kelautan. Vol 13(3) : 177-180
- [16] Bhadhury, P., & Wright , P. 2004. Exploitation of Marine Algae : Biogenic Compounds for Potential Antifouling Applications. Planta, 561–578.
- [17] Pérez, M., et al. 2009. Advances in Marine Antifouling Coatings and Technologies. Woodhead Publishing Ltd & CRC Press LLC, Cambridge, UK
- [18] Amin, M. 2017. Uji Ekstrak Daun Ketapang (*Terminalia catappa*) Sebagai Bahan Antifouling Alami Pada Plat Baja Di Perairan PT DOK Dan Perkapalan Surabaya. Surabaya: Institut Teknologi Sepuluh November.
- [19] Bordbar, S., Farooq A., dan Nazamid S. 2011. High-Value Components and Bioactives from Sea Cucumbers for Functional Foods²A Review [Marine Drugs Journal]. 1761-1805 hlm
- [20] Nimah, S., Widodo F.M, & Agus T. 2012. Uji Bioaktivitas Ekstrak Teripang Pasir (*Holothuria scabra*) Terhadap Bakteri *Pseudomonas aeruginosa* Dan *Bacillus cereus*. Jurnal Perikanan, 1.
- [21] Farouk, A. E. A., Ghouse, F. A. H. & Ridzwan, B. H. 2007. New Bacterial Species Isolated from Malaysian Sea Cucumbers with Optimized Secreted Antibacterial Activity, American Journal of Biochemistry and Biotechnology 3 (2), 60-65
- [22] Dwicahyani, T., Rianingsih , L., & Sumardianto. 2018. Uji Bioaktivitas Ekstrak Teripang Keling (*Holothuria atra*) Sebagai Antibakteri *Staphylococcus aureus* dan *Escherichia coli*. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan, 7.
- [23] Harborne, J. B. 1987. Metode Fitokimia: Penuntun Cara Modern MenganalisisTumbuhan. Terbitan Kedua, diterjemahkan oleh Padmawinata, K. & Sudiro, I. ITB Press, Bandung
- [24] Lubis, M., & Pujiyati, S. 2013. Pengaruh Aklimatisasi Kadar Garam Terhadap Nilai Kematian Dan Tingkah Laku Ikan Guppy (*Poecilia reculata*) Sebagai Pengganti Umpan Ikan Cakalang (*Katsuwonus pelamis*). Jurnal Teknologi Perikanan dan Kelautan, 123-129
- [25] Sastrosupadi, A. 2000. Rancangan Percobaan Praktis. Kanisus.Yogyakarta.276 hal
- [26] Arlyza, I. 2007. Bahan Aktif Dari Organisme Laut Sebagai Pengendali Biota Penempel. Jurnal Oseana, 32.
- [27] Masduki I, 1996. Efek Antibakteri Ekstrak Biji Pinang (*Areca catechu*) terhadap *S. aureus* dan *E. coli*. Cermin Dunia Kedokteran 109. pp. 4-21
- [28] Ajizah, Aulia. 2004. Sensitivitas *Salmonella typhimurium* terhadap ekstrak daun Psidium guajawa. Biosientiae Vol. 01: 31-38
- [29] Damayanti, E. dan T. B. Suparjana. 2007. Efek penghambatan beberapa fraksi ekstrak buah mengkudu terhadap *Shigella dysenteriae*. Prosiding Seminar Nasional Tehnik Kimia Keuangan. Fakultas Biologi Universitas Jenderal Soedirman. Yogyakarta
- [30] Cowan, M. 1999. Plant product as antimicrobial agents. Clinical Microbiology Reviews, 564 – 582
- [31] Bontjura S. 2015. Uji efek antibakteri ekstrak daun leilem (*Clerodendrum minahassae* l.) terhadap bakteri *streptococcus mutans*. Jurnal ilmiah Farmasi- Pharmacon.; 4 (4):96-101
- [32] Ahmed, Bahar. 2007. Chemistry Of Natural Products. New Delhi: Department of Pharmaceutical Chemistry Faculty of Science Jamia Hamdard.
- [33] Muchtadi, T.R. 1989. Teknologi Proses Pengolahan Pangan. Departemen Pendidikan dan Kebudayaan Direktorat Jenderal Pendidikan Tinggi Pusat Antar Universitas Pangan dan Gizi Institut Pertanian Bogor. Bogor.
- [34] Zhang, Y. S.; Yi, H. Y.; and Tang, H. F., 2006. Cytotoxic Sulfated Triterpene Glycosides from The Sea Cucumber *Pseudocolochirus violaceus*, Chemistry & Biodiversity, 3:807-817
- [35] Ma'ruf, & D. Pringgenies . 2012. Kajian Aktivitas Bioaktif Ekstrak Teripang Pasir (*Holothuria scabra*) Terhadap Jamur *Candida albicans*. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan, 1-8.
- [36] Hardiningtyas, S.D. 2009. Aktivitas Antibakteri Ekstrak Karang Lunak *Sarcophyton* sp. yang Difragmentasi dan Tidak Difragmentasi di Perairan Pulau Pramuka, Kepulauan Seribu. [Skripsi]. Institut Pertanian Bogor, Bogor, 67 hlm
- [37] Santi, I.W., O.K. Radjasa, dan I. Widowati. 2014. Potensi Rumput Laut *Sargassum duplicatum* Sebagai Sumber Senyawa Antifouling. Journal of Marine Research Vol. 3, No. 3: 274-284

- [38] Selvin J., & A.P Lipton. 2002. Development Of a Rapid "Mollusk Foot Adherence Bioassay" For Detecting Potent Antifouling Bioactive Compounds. *Current science*, 735-737.
- [39] KEPMEN LH No.59. 2004. Baku Mutu Bagi Biota Laut. Jakarta
- [40] Baragi. L.V., Anil. A.C. 2017. Influence of Elevated Temperature, pCO₂, and nutrients on larva-biofilm interaction: Elucidation with acorn barnacle, *Balanus amphitrite* Darwin (Cirripedia: Thoracica). *Estuarine, Coastal and Shelf Science*
- [41] Gemilang, W.A., Rahmawan, G.A., Wisha, U.J., 2017. Kualitas Perairan Teluk Ambon Dalam Berdasarkan Parameter Fisika dan Kimia pada Musim Peralihan I. *EnviroScientiae* 13(1) : 79-90
- [42] Kadim, M. K., Pasisingi, N., & Paramata, A. R. 2017. Kajian kualitas perairan Teluk Gorontalo dengan menggunakan metode STORET. *DEPIK Jurnal Ilmu-Ilmu Perairan, Pesisir dan Perikanan*, 6(3), 235-241
- [43] Idora, M. S., Ferry, M., Wan Nik, W., Jasnizat, S. 2015. Evaluation of tannin from *Rhizophora apiculata* as natural antifouling agents in epoxy paint for marine application. *Progress in Organic Coating* 81: 125-131
- [44] Selvin, J and Lipton, A.P. 2004. Antifouling activity of bioactive substances extracted from *Holothuria scabra*. *Hydrobiologia* 513, hh. 251-253
- [45] US EPA. 2002. Methods for Masuring The Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organism. Unites State: Environmental Protection Agency.
- [46] Wagner, J.G. 1993. Pharmacokinetics For The Pharmaceutical Scientist. Technomic Pub. Lancarter-Basel